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Certificate of Analysis

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HGPv1

Client: **Biohoney Limited** Contact: Terry Bone

> C/- Biohoney Limited 142 Collingwood Street

Nelson 7010

Lab No: **Date Received: Date Reported:**

05-Nov-2021 08-Nov-2021

2757496

96232

Quote No: Order No:

Client Reference:

Submitted By: Terry Bone

Sample Type: Honey						
	Sample Name:	2104RM400				
	Lab Number:	2757496.1				
Manuka Honey Analysis						
Dihydroxyacetone (DHA)	mg/kg	711	-	-	-	-
5-Hydroxymethylfurfural (HMF)	mg/kg	28.1	-	-	-	-
Methylglyoxal (MGO)	mg/kg	429	-	-	-	-
Non Peroxide Activity (NPA)*	% Phenol Equivalent	13.4	-	-	-	-

Summary of Methods

The following table(s) gives a brief description of the methods used to conduct the analyses for this job. The detection limits given below are those attainable in a relatively simple matrix. Detection limits may be higher for individual samples should insufficient sample be available, or if the matrix requires that dilutions be performed during analysis. A detection limit range indicates the lowest and highest detection limits in the associated suite of analytes. A full listing of compounds and detection limits are available from the laboratory upon request Unless otherwise indicated, analyses were performed at Hill Laboratories, 28 Duke Street, Frankton, Hamilton 3204.

Sample Type: Honey							
Test	Method Description	Default Detection Limit	Sample No				
3-in-1 Honey Method	Aqueous extraction, derivatisation. Analysis by uHPLC / UV-Vis (dihydroxyacetone, 5-hydroxymethylfurfural, methylglyoxal). Inhouse.	-	1				
Non Peroxide Activity (NPA)*	NPA is calculated from methylglyoxal using a correlation curve based on published data for NPA and the primary active ingredient, methylglyoxal. (1,2). (1) Isolation by HPLC and characterisation of the bioactive fraction of New Zealand manuka (Leptospermum scoparium) honey. C. J. Adams, et al. Carbohydrate Research 343 (2008) 651-659. (2) Corrigendum to "Isolation by HPLC and characterization of the bioactive fraction of New Zealand manuka (Leptospermum scoparium) honey" [Carbohydr. Res. 343 (2008) 651]. C. J. Adams, et al. Carbohydrate Research 344 (2009) 2609.	1.0 % Phenol Equivalent	1				

These samples were collected by yourselves (or your agent) and analysed as received at the laboratory.

Testing was completed on 08-Nov-2021. For completion dates of individual analyses please contact the laboratory.

Samples are held at the laboratory after reporting for a length of time based on the stability of the samples and analytes being tested (considering any preservation used), and the storage space available. Once the storage period is completed, the samples are discarded unless otherwise agreed with the customer. Extended storage times may incur additional charges.

This certificate of analysis must not be reproduced, except in full, without the written consent of the signatory.

Helen McGowan BSc (Tech)

Operations Support - Food & Bioanalytical



